



# Reproductive capacity of *varroa destructor* in four different honey bee subspecies

Richard Odemer<sup>1</sup>

University of Hohenheim, Apicultural State Institute, August-von-Hartmann-Str. 13, 70593 Stuttgart, Germany

## ARTICLE INFO

### Article history:

Received 17 October 2018

Revised 27 August 2019

Accepted 1 September 2019

Available online 9 September 2019

### Keywords:

Varroa mite

Reproduction

Offspring

*Apis mellifera* subspecies

Fecundity

Fertility

## ABSTRACT

Varroa tolerance as a consequence of host immunity may contribute substantially to reduce worldwide colony declines. Therefore, special breeding programs were established and varroa surviving populations investigated to understand mechanisms behind this adaptation. The aim of this study was to investigate the reproductive capacity in the three most common subspecies of the European honey bee (*Carnica*, *Mellifera*, *Ligustica*) and the F2 generation of a varroa surviving population, to identify if managed host populations possibly have adapted over time already. Both, singly infested drone and worker brood were assessed to determine fertility and fecundity of varroa foundresses in their respective group. We found neither parameter to be significantly different within the four subspecies, demonstrating that no adaptations have occurred in terms of the reproductive success of *Varroa destructor*. In all groups mother mites reproduce equally successful and are potentially able to cause detrimental damage to their host when not being treated sufficiently. The data further suggests that a population once varroa tolerant does not necessarily inherit this trait to following generations after the F1, which could be of particular interest when selecting populations for resistance breeding. Reasons and consequences are discussed.

© 2019 Production and hosting by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 1. Introduction

Varroosis is known to be the most serious threat for European honey bees across the globe (Rosenkranz et al., 2010). A key for the mite's success lies in their ability to perfectly adapt to host conditions, including the reproduction in worker brood. Even though reproductive capacity of *V. destructor* seems equally high in both, drone and worker brood, a distinctive amount of mites fail to reproduce even though they are not infertile (de Ruijter, 1987). The conditions however, under which mite foundresses remain "temporary sterile" cannot yet be explained (Garrido and Rosenkranz, 2003) but is discussed to be a host-specific tolerance trait against the mite (Rosenkranz and Engels, 1994). Host stages in which mites are able to reproduce vary between drone and worker brood and reproduction is only possible within a narrow

time frame, indicating a particularly sensitive process (Frey et al., 2013). Interestingly, Xie et al. (2016) revealed that mother mites are able to choose nurse bees over foragers and newly emerged bees as their optimal host in the phoretic phase, not only enabling them to quickly infest new brood cells (Donzé et al., 1998), but also providing the best possible nutritional conditions to produce a larger amount of progeny. Subsequently, the varroa population per colony can increase up to ten times in only one short beekeeping season (Sokół et al., 2019) which overall demonstrates a high degree of adaptation.

Reports from surviving populations have increased over the last decade, suggesting a rapid host adaptation more or less simultaneously (Oddie et al., 2018). Besides a specific varroa mite targeted hygienic behavior (VSH = varroa sensitive hygiene) (Panziera et al., 2017), reduced mite reproduction is considered to be one key advantage for colony survival by means of natural selection (Locke et al., 2012). Almost exclusively, such traits have been investigated and documented for resistant honey bee populations (Locke, 2016a) but have probably been neglected for more common subspecies. To date, investigations on the mites' reproductive success have focused on exotic bee subspecies such as *A.m. syriaca* (Alattal and Rosenkranz, 2006) or the Africanized honey bee (Garrido et al., 2003). Little or nothing is known about the adaptation potential of subspecies which are native to most parts of

E-mail address: [richard.odemer@uni-hohenheim.de](mailto:richard.odemer@uni-hohenheim.de)

<sup>1</sup> Present address: Julius Kuehn-Institute (JKI), Federal Research Centre for Cultivated Plants, Institute for Bee Protection, 38104 Brunswick, Germany.

Peer review under responsibility of King Saud University.



Production and hosting by Elsevier

Europe. To close this knowledge gap and ascertain both, fertility and fecundity as a consequence of the reproductive capacity of *V. destructor*, we have compared the three most common subspecies of the European honey bee, i.e. *Apis mellifera carnica* (branch-M, western Europe), *A. m. mellifera* (branch-M, northern Europe) and *A. m. ligustica* (branch-C, southeastern Europe) representing at least two different evolutionary branches, corresponding to distinct geographic areas in Europe to cover a wide range of adaptation potential (Bouga et al., 2011). In addition, the F2 generation of a varroa surviving population descending from the “Bond Project” on Gotland (Fries et al., 2006) was evaluated, to identify if managed host populations possibly have adapted over time already despite systematic control measures.

## 2. Materials & methods

### 2.1. Bee colonies and subspecies

A total of 22 honey bee colonies (*A. mellifera* L.) were investigated during summer season from May to August. We focused on subspecies originating in Europe such as the Carniolan bee *A. m. carnica* (n = 5, originated from our local Hohenheim breeding line), the European dark bee *A. m. mellifera* (n = 7, originated from a pure-breeder in Freiburg, Germany), the Italian bee *A. m. ligustica* (n = 5, originated from a pure-breeder in Alsace, France) and a F2 generation of mite surviving bees from the “Bond Project” descending from the Swedish island of Gotland “Gotland/F2” (n = 5). To provide a sufficient amount of drone pupa, one to two drone-frames were placed at the edge of the brood nest of each colony. All experimental hives were either successfully overwintered from the past season (Carnica, Go/F2, Mellifera) or freshly created by re-queening established colonies (Ligustica). They were kept and maintained without varroa treatment in the current season at our local apiary near the Apicultural State Institute in Stuttgart, Germany.

### 2.2. Mite reproduction

The reproductive capacity of the foundress mite is specified as success to generate at least one viable daughter before the host pupa hatches (fertility). In contrast, mother mites that lay no or only a single egg, have no males or are delayed in egg-laying respective to host-development will fail to produce viable offspring for the following mite generations. Further, the number of progeny per mite (fecundity) serves as measure for a possible host adaptation representing a reduced reproductive capacity in terms of an increased survivability of the colony.

To increase comparability of our results, all experiments were performed according to the methods described in Locke and Fries (2011). In brief, worker and drone pupae in stage Pd and older, but before eclosion, were examined (see Fig. 1). At least 30 cells per colony were carefully investigated where possible and mite

infestation was documented. Only cells with a single foundress were considered, cell content and mites attached to the pupa were accurately removed and subsequently observed under a stereomicroscope (Zeiss Stemi, 2000-CS). Varroa mites were able to naturally infest drone and worker brood in all colonies, no additional mites were inserted.

### 2.3. Data evaluation

Mite reproduction and fecundity data were first tested for variance homogeneity and normal distribution with Levene’s and Shapiro-Wilk test and verified for both datasets, respectively. A generalized linear model was applied to both sets followed by a comparison of the least-squares means and a *P* value adjustment (Tukey method i.e. Tukey’s HSD test). For all tests RStudio (R Core Team, 2018) and significance level of  $\alpha = 0.05$  was used.

## 3. Results

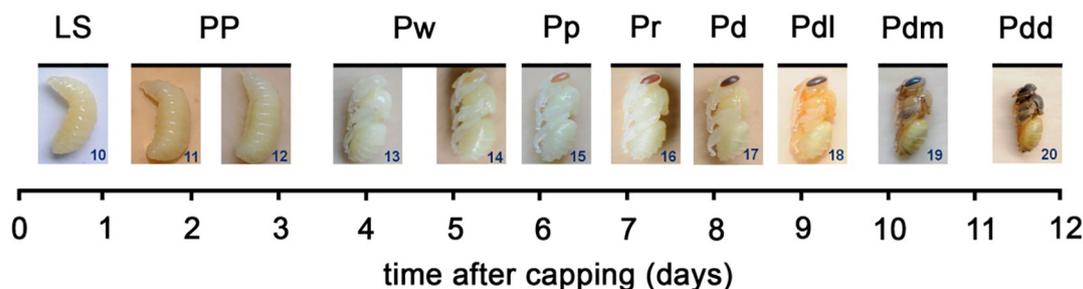
Different parameters of varroa mite reproduction in four different honey bee subspecies are presented in Table 1. A total of n = 3104 drone and n = 2526 worker brood cells were evaluated, including empty and multiply infested cells. We did not find significant differences for the overall reproductive capacity (fertility) in the four groups. Neither in worker brood (*df* = 10: *F* = 2.26; *P* = 0.144) nor in drone brood (*df* = 15: *F* = 2.51; *P* = 0.098). A similar outcome was observed for the average number of offspring per foundress (fecundity). Both, progeny found in worker brood (*df* = 10: *F* = 2.84; *P* = 0.092) and in drone brood (*df* = 10: *F* = 2.32; *P* = 0.873) were at the same level.

Due to an increased infestation rate which resulted in a high ratio of multiply infested cells in the drone brood of all four subspecies, it was not possible to evaluate drone pupa in stage Pd and older as previously described. To compare fecundity regardless these circumstances, we had to consider earlier developmental stages beginning already at Pw (Fig. 1) providing a sufficient amount of singly infested cells. This is why the average number of offspring is relatively low when compared to worker brood.

For the number of cells in Ligustica drone brood it needs to be mentioned that due to the late re-queening of experimental colonies (mid July) it was not possible to obtain a sufficient amount of singly infested cells. Hence, we only used 10 cells per colony on average, this should be considered when interpreting the results.

## 4. Discussion

Here, we studied the reproductive capacity of three commonly managed honey bee subspecies and the F2 generation of a varroa surviving population originated from the “Bond Project” (Fries



**Fig. 1.** Classification of pupal stages relative to ontogenetic worker development (after Rembold et al., 1980, graphically modified after Wang et al., 2015). Abbreviations: LS = 5th larval instar after sealing; PP = prepupa; P = pupa (w = white eyes; p = pink eyes; r = red eyes; d = dark brown eyes; dl = dark brown eyes, light pigmented thorax; dm = dark brown eyes, medium colored thorax; dd = dark brown eyes, dark thorax).

**Table 1**Comparison of the reproductive capacity (mean fertility and fecundity  $\pm$  standard error) of mother mites produced in singly infested drone and worker brood cells]

	Carnica	Mellifera	Ligustica	Gotland/F2	
<b>Drones</b>					
Total No. of cells (n)	68	179	51 <sup>b</sup>	141	
Mean fertility ( $\pm$ SE)	79% ( $\pm$ 8.4)	83% ( $\pm$ 5.5)	59% ( $\pm$ 7.3)	79% ( $\pm$ 6.5)	ns
Mean fecundity ( $\pm$ SE) <sup>a</sup>	2.7 ( $\pm$ 0.5)	2.7 ( $\pm$ 0.3)	2.2 ( $\pm$ 0.6)	2.6 ( $\pm$ 0.2)	ns
<b>Workers</b>					
Total No. of cells (n)	90	91	120	120	
Mean fertility ( $\pm$ SE)	82% ( $\pm$ 6.1)	89% ( $\pm$ 6.1)	96% ( $\pm$ 5.2)	78% ( $\pm$ 5.2)	ns
Mean fecundity ( $\pm$ SE)	3.3 ( $\pm$ 0.3)	3.4 ( $\pm$ 0.3)	4.1 ( $\pm$ 0.4)	3.3 ( $\pm$ 0.2)	ns

ns: not significant ( $P > 0.05$ ).<sup>a</sup> Earlier developmental stages beginning already at Pw had to be considered for the drone brood.<sup>b</sup> Not representative, due to the low amount of singly infested cells (10 cells per colony on average).

et al., 2006). When compared to former data, the fertility of varroa foundresses in worker brood did not change significantly during the past three decades and has levelled off between 80 and 90 % (Thrybom and Fries 1991; Corrêa-Marques et al., 2003; Garrido et al., 2003; Alattal and Rosenkranz, 2006; Locke et al., 2012; Alattal et al., 2017). This trend is corroborated by our data and most likely similar for drone brood.

Drone frames that we have investigated here were highly infested already in early summer, not least because some colonies remained untreated in the former season at our experimental apiary but also because the mite's preference to infest drone cells is approximately eight times higher when compared to worker brood (Fuchs, 1990; Santillán-Galicia et al., 2002). In addition, the time frame which is attractive to enter cells for infestation is approximately twice as long in drone brood (Calderone et al., 2002), being one reason for this preference. Under these circumstances it was not surprising that we found many multiply infested drone cells and it became a challenge to locate cells containing only one foundress for our evaluation. Ligustica queens arrived after summer solstice very late in the season and besides that, a very high mite infestation in drones was the reason that we were not able to collect a sufficient amount of singly infested cells.

Moreover, our data confirms that there is no large selection pressure favoring reduced mite reproduction in both, drones and workers, at least not under intensively managed conditions. For the three common subspecies this is not remarkable as host adaptations are most often reported as a means of natural selection (Seeley, 2007; Locke et al., 2012; Oddie et al., 2017). For the F2 generation of the surviving population from Gotland however, we had expected a different outcome. The Gotland bees have developed an apparent reduced mite reproductive success trait that is either inheritable from paternal, maternal or both sides in the F1 generation (Locke, 2016b). Our results provide evidence that this trait seems to fade out by further generational change, once more making the colonies susceptible to *Varroosis*.

Although we did not find significant differences in the fertility and fecundity of varroa females between surviving F2 and common honey bee subspecies, we are still convinced that the varroa reproductive capacity represents a crucial and probably the only parameter for the future selection of varroa resistance on the individual level. One reason is that we confirmed that about 85% of the “temporary sterile mites” were again fertile if re-introduced into freshly sealed brood cells (Weller, 2008). Hence, the occurrence of “temporary sterile mites” seems to be rather a trait of the host than a trait of the parasite and, therefore, offers possibilities for selection.

## 5. Conclusion

Frequent reports have shown that apart from the most common managed honey bee subspecies there are populations demonstrat-

ing increased mite susceptibility and great variance in mite reproductive capacity (de Guzman et al., 2008; Locke, 2016a; Nganso et al., 2018). This reflects an encouraging potential to establish varroa resistance in European *A. mellifera* populations (Büchler et al., 2010). However, resistance mechanisms are complex which is why further research is necessary to understand host-adaptation and mite reproduction in greater detail.

## Acknowledgements

I would like to thank Peter Rosenkranz and Eva Frey for providing help and infrastructural support to conduct the experiments.

This research received funding from the BEE SHOP (BEes in Europe and Sustainable HONEY Production), grant contract no.: PL 022568.

The funding source had no influence on study design; the collection, analysis and interpretation of data; the writing of the report; and the decision to submit the article for publication in this journal.

## Declaration of Competing Interest

None.

## Ethical approval

This article does not contain any studies with human participants or animals performed by any of the authors.

## References

- Alattal Y, Rosenkranz P, Zebitz CPW (2006) Reproduction of *Varroa destructor* in sealed worker bee brood cells of *Apis mellifera carnica* and *Apis mellifera syriaca* in Jordan. *Mitt Dtsch Ges Allg Angew Ent* 315–320.
- Alattal, Y., AlGhamdi, A., Single, A., Ansari, M.J., Alkathiri, H., 2017. Fertility and reproductive rate of *Varroa mite, Varroa destructor*, in native and exotic honeybee, *Apis mellifera L.*, colonies under Saudi Arabia conditions. *Saudi J. Biol. Sci.* 24, 992–995. <https://doi.org/10.1016/j.sjbs.2016.12.018>.
- Bouga, M., Alaux, C., Bienkowska, M., Büchler, R., Carreck, N.L., Cauia, E., Wilde, J., 2011. A review of methods for discrimination of honey bee populations as applied to European beekeeping. *J. Apicult. Res.* 50 (1), 51–84. <https://doi.org/10.3896/IBRA.1.50.1.06>.
- Büchler, R., Berg, S., Le Conte, Y., 2010. Breeding for resistance to *Varroa destructor* in Europe. *Apidologie* 41, 393–408. <https://doi.org/10.1051/apido/2010011>.
- Calderone, N., Lin, S., Kuenen, L., 2002. Differential infestation of honey bee, *Apis mellifera*, worker and queen brood by the parasitic mite *Varroa destructor*. *Apidologie* 33, 389–398. <https://doi.org/10.1051/apido:2002024>.
- Corrêa-Marques, M.H., Medina, L.M., Martin, S.J., De Jong, D., 2003. Comparing data on the reproduction of *Varroa destructor*. *Genet. Mol. Res.* 2, 1–6.
- de Guzman, L.I., Rinderer, T.E., Frake, A.M., 2008. Comparative reproduction of *Varroa destructor* in different types of Russian and Italian honey bee combs. *Exp. Appl. Acarol.* 44, 227–238. <https://doi.org/10.1007/s10493-008-9142-1>.
- de Ruijter, A., 1987. Reproduction of varroa jacobsoni during successive brood cycles of the honeybee. *Apidologie* 18, 321–326. <https://doi.org/10.1051/apido:19870403>.

- Donzé, G., Fluri, P., Imdorf, A., 1998. Hochorganisierte Leben auf kleinem Raum: Die Fortpflanzung der Varroa-Milben in den verdeckelten Brutzellen der Bienenvölker. *Schweizerische Bienen-Zeitung* 121, 26–33.
- Fries, I., Imdorf, A., Rosenkranz, P., 2006. Survival of mite infested (*Varroa destructor*) honey bee (*Apis mellifera*) colonies in a Nordic climate. *Apidologie* 37, 564–570. <https://doi.org/10.1051/apido>.
- Frey, E., Odemer, R., Blum, T., Rosenkranz, P., 2013. Activation and interruption of the reproduction of *Varroa destructor* is triggered by host signals (*Apis mellifera*). *Journal of Invertebrate Pathology* 113 (1), 56–62. <https://doi.org/10.1016/j.jip.2013.01.007>.
- Fuchs, S., 1990. Preference for drone brood cells by *Varroa jacobsoni* Oud in colonies of *Apis mellifera carnica*. *Apidologie* 21, 193–199. <https://doi.org/10.1051/apido:19900304>.
- Garrido, C., Rosenkranz, P., 2003. The reproductive program of female *Varroa destructor* mites is triggered by its host, *Apis mellifera*. *Exp. Appl. Acarol.* 31, 269–273. <https://doi.org/10.1023/B:APPA.0000010386.10686.9f>.
- Garrido, C., Rosenkranz, P., Paxton, R.J., Gonçalves, L.S., 2003. Temporal changes in *Varroa destructor* fertility and haplotype in Brazil. *Apidologie* 34 (6), 535–541. <https://doi.org/10.1051/apido:2003041>.
- Gullan, P.J., Cranston, P.S., 2014. *The Insects: An Outline of Entomology*. John Wiley & Sons Blackwell, Oxford.
- Locke, B., 2016a. Natural *Varroa* mite-surviving *Apis mellifera* honeybee populations. *Apidologie* 47, 467–482. <https://doi.org/10.1007/s13592-015-0412-8>.
- Locke, B., 2016b. Inheritance of reduced *Varroa* mite reproductive success in reciprocal crosses of mite-resistant and mite-susceptible honey bees (*Apis mellifera*). *Apidologie* 47, 583–588. <https://doi.org/10.1007/s13592-015-0403-9>.
- Locke, B., Fries, I., 2011. Characteristics of honey bee colonies (*Apis mellifera*) in Sweden surviving *Varroa destructor* infestation. *Apidologie* 42, 533–542. <https://doi.org/10.1007/s13592-011-0029-5>.
- Locke, B., Le Conte, Y., Crauser, D., Fries, I., 2012. Host adaptations reduce the reproductive success of *Varroa destructor* in two distinct European honey bee populations. *Ecol. Evol.* 2, 1144–1150. <https://doi.org/10.1002/ece3.248>.
- Nganso, B.T., Fombong, A.T., Yusuf, A.A., Pirk, C.W.W., Stuhl, C., Torto, B., 2018. Low fertility, fecundity and numbers of mated female offspring explain the lower reproductive success of the parasitic mite *Varroa destructor* in African honeybees. *Parasitology* 1–7. <https://doi.org/10.1017/S0031182018000616>.
- Oddie, M., Dahle, B., Neumann, P., 2017. Norwegian honey bees surviving *Varroa destructor* mite infestations by means of natural selection. *PeerJ* 5, <https://doi.org/10.7717/peerj.3956> e3956.
- Oddie, M., Büchler, R., Dahle, B., Kovacic, M., Le Conte, Y., Locke, B., de Miranda, J.R., Mondet, F., Neumann, P., 2018. Rapid parallel evolution overcomes global honey bee parasite. *Sci. Rep.* 8, 7704. <https://doi.org/10.1038/s41598-018-26001-7>.
- Panziera, D., van Langevelde, F., Blacquièrre, T., 2017. *Varroa* sensitive hygiene contributes to naturally selected *Varroa* resistance in honey bees. *J. Apic. Res.* 56, 635–642. <https://doi.org/10.1080/00218839.2017.1351860>.
- Core Team, R., 2018. *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Rembold, H., Kremer, J.-P., Ulrich, G.M., 1980. Characterization of postembryonic developmental stages of the female castes of the Honey Bee, *Apis mellifera* L. *Apidologie* 11, 29–38. <https://doi.org/10.1051/apido:19800104>.
- Rosenkranz, P., Aumeier, P., Ziegelmann, B., 2010. Biology and control of *Varroa destructor*. *J. Invertebr. Pathol.* 103 (Suppl), S96–S119. <https://doi.org/10.1016/j.jip.2009.07.016>.
- Rosenkranz, P., Engels, W., 1994. Infertility of *Varroa jacobsoni* females after invasion into *Apis mellifera* worker brood as a tolerance factor against varroaosis. *Apidologie* 25 (4), 402–411. <https://doi.org/10.1051/apido:19940407>.
- Santillán-Galicia, M., Otero-Colina, G., Romero-Vera, C., Cibrián-Tovar, J., 2002. *Varroa destructor* (Acari: Varroidae) infestation in queen, worker, and drone brood of *Apis mellifera* (Hymenoptera: Apidae). *Can. Entomol.* 134 (3), 381–390. <https://doi.org/10.4039/Ent134381-3>.
- Seeley, T., 2007. Honey bees of the Arnot Forest: a population of feral colonies persisting with *Varroa destructor* in the northeastern United States. *Apidologie* 38, 19–29. <https://doi.org/10.1051/apido:2006055>.
- Sokół, R., Gałęcki, R., Michalczyk, M., 2019. Controlled Infestation of Honeybee Colonies with *Varroa Destructor* Females. *J. Apicult. Sci.* 63 (1), 149–155. <https://doi.org/10.2478/jas-2019-0009>.
- Thrybom, B., Fries, I., 1991. Development of infestations by *Varroa jacobsoni* in hybrid colonies of *Apis mellifera monticola* and *Apis mellifera ligustica*. *J. Apicult. Sci.* 30 (3–4), 151–155. <https://doi.org/10.1080/00218839.1991.11101250>.
- Wang, Y., Ma, L.-T., Xu, B.-H., 2015. Diversity in life history of queen and worker honey bees, *Apis mellifera* L. *J. Asia Pac. Entomol.* 18, 145–149. <https://doi.org/10.1016/j.aspen.2014.11.005>.
- Weller, S. (2008). Populationsdynamik der parasitischen Bienenmilbe *Varroa destructor* in vorselektierten Bienenvölkern (*A. mellifera* L.) unter besonderer Berücksichtigung der Reproduktion. Diploma Thesis, Apicultural State Institute, Stuttgart-Hohenheim.
- Xie, X., Huang, Z.Y., Zeng, Z., 2016. Why do *Varroa* mites prefer nurse bees? *Sci. Rep.* 6, 28228. <https://doi.org/10.1038/srep28228>.